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### RESEARCH ARTICLE

#### GREEN SYNTHESIS, CYTOTOXICITY AND UTILIZATION OF CARBOXYMETHYLCHITOSAN-STABILIZED GOLD NANOPARTICLES.

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#### Abstract

Gold nanoparticles (AuNPs) was prepared via simple and green method by using polysaccharides as reducing and stabilizing agents at the same time. Hiren we used carboxymethylchitosan (CMCS) as a reducing agent for gold nanoparticle as well as capping agent. CMCS prepared based on our previous method by reacting chitosan with monochloroacetic acid in alkaline medium. AuNPs were prepared by using different concentrations of carboxymethylchitosan (0.2% w/v, 0.5% w/v and 1% w/v) at 100 °C for 1 hour. CMCS was characterized by using nitrogen content, carboxyl content and FTIR spectra. AuNPswas characterized by using UV spectrophotometry and TEM images. Finally, the cytotoxicity of the prepared AuNPs were evaluated using cell viability assay from MMT and IC<sub>50</sub> values compared with AuNPs prepared by chemical methods. The results shows that AuNPs have normal distributed with 15-25 nm particle size and its cytotoxicity was lowered when prepared by this green method and can use GNPs safely in contact medical treatment with skin.

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#### Introduction:-

Metal nanoparticles preparation represents major area in nanoscale science and engineering to give unusual physical and chemical properties specially its catalytic activity, novel electronic and magnetic properties and their potential applications in bio nanotechnology[1, 2].

Metal nanoparticles generally prepared by chemical reduction of metal salts with chemical reducing agents such as citric acids, borohydrides, or other organic compounds[1, 3-5].These reducing agents cause cytotoxicity towards biological hazards. Green chemistry used to minimize or eliminate the waste and implement sustainable process [6].So that biological method used for prepare AuNPs.

Raveendran et al was the first team work used the green concept to prepare silver nanoparticles by using glucose as reducing agent and starch as capping agent [7]. Nanoparticles preparation via green method was evaluated from three aspects: solvent, reducing agent and capping agent[8].

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Chitosan as inexpensive material with renewable sources used in several applications especially in cosmetics, pharmaceuticals, food and biotechnology [9, 10]. In the preparation of gold nanoparticles by using chitosan biopolymer, the  $\text{NH}_2$  groups used to stabilize gold nanoparticles [11, 12]. However precise control needed to avoid nanoparticles aggregations [11, 13]. Due to poor solubility of chitosan. Few papers found in the literature dealing with the direct application of chitosan and AuNPs nanocomposites.

Water-soluble chitosan derivative, O-carboxymethylchitosan (CMCS), not only has good solubility in water but also has unique properties (chemical, physical and biological) e.g. high viscosity, biodegradability, biocompatibility and low cytotoxicity. Herein chemical modification perform only on OH groups to form  $\text{OCH}_2\text{COOH}$ , which retain  $\text{NH}_2$  groups accessible for reaction i.e., O-CMCS has both COOH and  $\text{NH}_2$  groups used to stabilize AuNPs so that O-CMCS used as capping agent as well as reducing agent [14].

In the present study, we used simple and green method for gold nanoparticles preparation using carboxymethylchitosan as reducing agent and capping agent at the same time. No other chemical substances needed for the reduction process. We used aqueous solution in this process to avoid environmental hazards. UV spectrophotometry and TEM imaging used to characterize the prepared AuNPs. Finally, we evaluated the cytotoxicity of these nanoparticles compared with the AUGPs prepared by common chemical reduction.

## **Experimental:-**

### **Materials:-**

Chitosan (CS) (Aldrich, viscosity 1860cps, degree of deacetylation 79.0%). Sodium hydroxide (Modern Lab chemicals), monochloroacetic (Fluka), are used without further purification. Auric-chloride ( $\text{HAuCl}_4$ ) purchased from Aldrich and used without further purification, USA. All other chemicals and reagents were of analytical grade, and were used without further purification.

### **Methods:-**

#### **Preparation of Carboxymethylchitosan:-**

The carboxymethylation of chitosan (CMCS) was prepared as reported in our previous work [15, 16].

#### **Preparation of Gold Nanoparticles (AuNPs):-**

Gold nanoparticles were prepared by reduction of sodium citrate method as mentioned where [17, 18] with some slight modifications as follow: gold (III) chloride stock solution (1%) used to prepare 15mM, 20mM and 25mM respectively, followed by heating to  $95^\circ\text{C}$  under stirring on magnetic stirrer with heater. To this boiling solution add tri sodium citrate (1.5% w/v), and continue stirring until give red colour. Then we stored this solution at  $4^\circ\text{C}$  to be ready for use.

#### **Finishing of Fabrics with Gold nanoparticles:-**

The prepared gold nanoparticles (AuNPs) were applied on washed and dried fabrics using pad-dry-cure method.  $30 \times 30$  cm of fabrics were immersed in the gold nanoparticles (AuNPs) (0.005 -0.5 g/ml) solution containing acrylate binder (1%) for 30 min., and then it was passed through a padding mangle with 100% wet pick-up for all of the treatments. Then the fabrics were dried at  $80^\circ\text{C}$  for 5 min., followed by thermo-fixation for at  $140^\circ\text{C}$  for 3 min. Finally, samples washed and dried to be ready for characterization and antibacterial evaluation.

#### **Characterizations of Gold Nanoparticles (AuNPs):-**

- Fourier transform infrared (FT-IR) spectra of the samples were recorded by using an FT-IR spectrophotometer (Nexus 670, Nicolet, USA) in the region of  $4000\text{--}400\text{cm}^{-1}$  with spectra resolution of  $4\text{ cm}^{-1}$ .
- UV-vis spectroscopy of AuNPs were record on Shimadzu (UV-2450) to confirm the presence of AuNPs in the reaction medium at range 510–560 nm.
- Shape and size of gold nanoparticles (AuNPs) were investigated using JEOL, JXA-840 electron probe microanalyzer, Japan.
- The UV-protection factor (UPF) demonstrates the ratio of sunburn-causing UV measured without and with the protection of the fabric. The UPF of untreated and finished fabric samples (size  $3\text{ cm} \times 3\text{ cm}$ ) was determined according to the Australian/New Zealand standard (AS/NZS 4399-1996: Sun protective clothing-Evaluation and classification) using UV-Shimadzu 3101 PC spectrophotometer at wavelength of 280–390 nm, which includes the UVB (280–320 nm) and the UVA (320–400 nm) according to the following equation:

$$UPF_i = \frac{\sum_{\lambda=280}^{400} E_{\lambda} \times S_{\lambda} \times \Delta\lambda}{\sum_{\lambda=280}^{400} E_{\lambda} \times S_{\lambda} \times T_{\lambda} \times \Delta\lambda}$$

where:  $E_{\lambda}$  = relative erythral spectral effectiveness,  $S_{\lambda}$  = solar spectral irradiance,  $T_{\lambda}$  = average spectral transmission of the specimen, and  $\Delta\lambda$  = measured wavelength interval (nm) Regarding UV –protection categories, fabrics are classified to good, very good, and excellent if their UPF values range from 15 to 24, 25 to 39, and above 40 (40+) respectively.

- SEM and EDX of the treated fabrics was studied using a scanning electron probe micro analyzer (type JXA 840A)–Japan. Surface morphologies were imaged at different magnifications, using 30kV accelerating voltage.

The tests was carried out at the Central unit for analysis and scientific services at National Research Center.

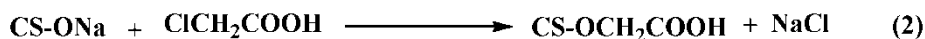
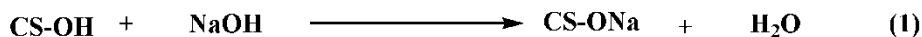
### Evaluation of cytotoxicity of AuNPs:

Cytotoxicity of the prepared AuNPs on A-549 cells were evaluated via cell viability test using MMT method (3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide) and determination of values of  $IC_{50}$  [19, 20].

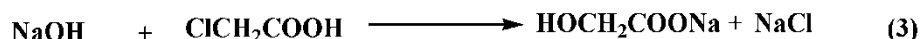
### Results and Discussion:-

#### Preparation of Carboxymethylchitosan:-

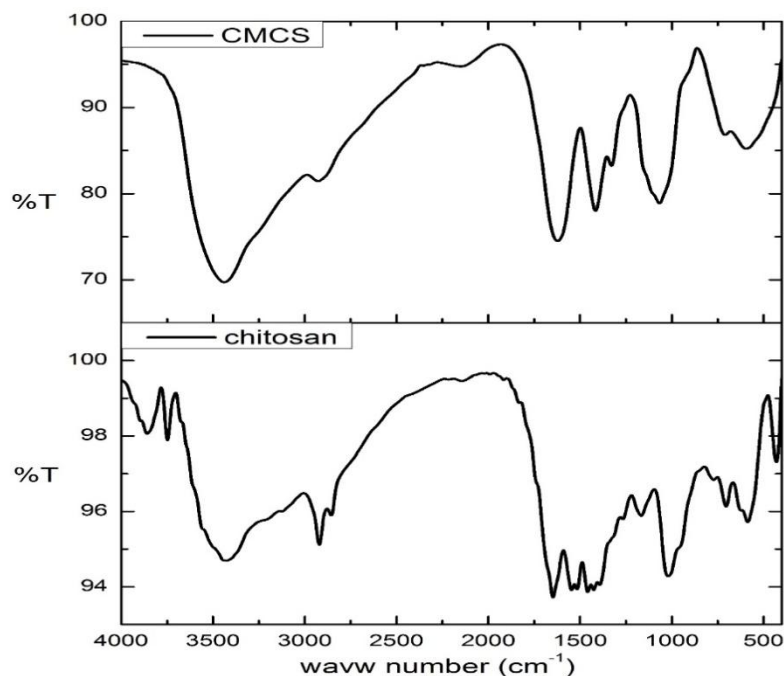
The carboxymethylation of chitosan proceeds by two step consecutive reaction and is accompanied by an undesired side reaction. In the main reaction, the sodium hydroxide reacts first with the hydroxyl groups of chitosan to give alkali chitosan. The carboxymethyl groups are then formed in a SN2 reaction between the alkali chitosan and monochloroacetic acid (MCAA). The main reaction is given by:



The side reaction takes place and results in the formation of sodium glycolate from MCAA and sodium hydroxide.



The FTIR spectra of the prepared CMCSs is shown in Figure 1. In IR spectrum, the wide band at  $3420\text{ cm}^{-1}$  corresponds to the axial stretching of the O–H and N–H bonds. The peaks at  $2927\text{ cm}^{-1}$  and  $1639\text{ cm}^{-1}$  are attributed to the axial stretching of the C–H bonds and the symmetric stretching vibration of C=O in the –COOH groups, respectively. The latter peak, together with the peak at  $1420\text{ cm}^{-1}$ , which arose from the asymmetric stretching vibration of the –COO<sup>−</sup> group, confirm the substitution of carboxymethyl groups onto the chitosan chain. Two bands at  $1528$  and  $1513\text{ cm}^{-1}$  assigned to NH<sub>3</sub><sup>+</sup>, indicate that the carboxymethylation occurred at OH positions. The peaks at  $1413$  and  $1377\text{ cm}^{-1}$  are related to the symmetric angular deformation of C–H bonds and C–N stretching vibrations (amide III band), respectively. The peak at  $1377\text{ cm}^{-1}$  did not increase significantly in the spectra of the CMCS, compared to the chitosan spectrum, which indicates that a significant amount of N-carboxymethylation did not take place. The stretching vibration of C–O in the CH<sub>2</sub>COOH group gives rise to the peak at  $1207\text{ cm}^{-1}$ . Peaks located in the range of  $1175$ – $878\text{ cm}^{-1}$  are the result of vibrations of C–O and C–O–C and some other bonds that comprise the polysaccharide chain [21].

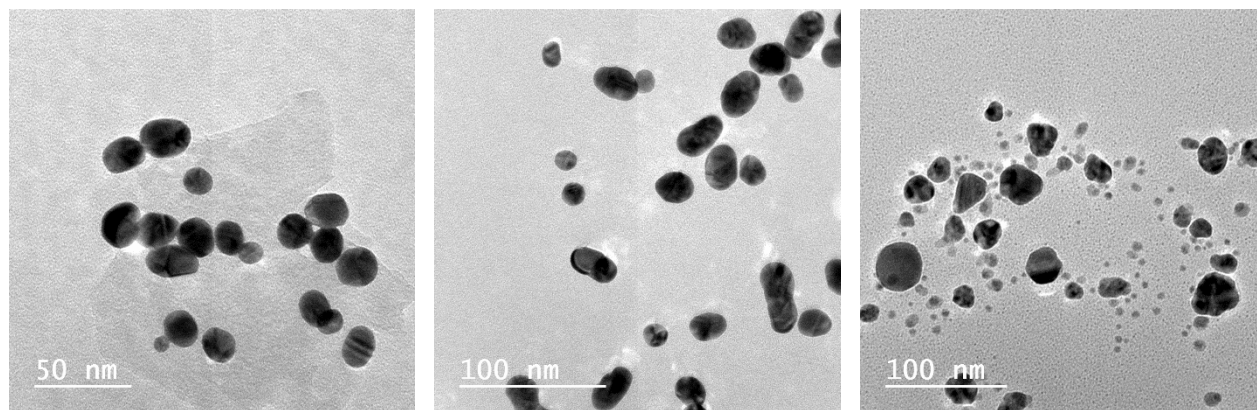


**Figure 1:**-FTIR spectrum of chitosan (CS) and CMCS prepared by reaction of 5gmchitosan with 2.5M MCAA in the presence of 50% NaOH within 3hrs at60°C.

#### Characterization of the CMCS–AuNPs nanocomposite:-

Carboxymethylchitosan play an important role in the preparation of nanoparticles. Amino groups have been used as metal nanoparticles stabilizer [14, 22]. So that we suggest that free amino groups in O-CMCS could bind with gold nanoparticles to stabilize it. When chitosan used instead of O-CMCS to stabilize AuNPs aggregation of nanoparticles occurs due to chitosan insoluble in both neutral and alkaline medium [11, 14].

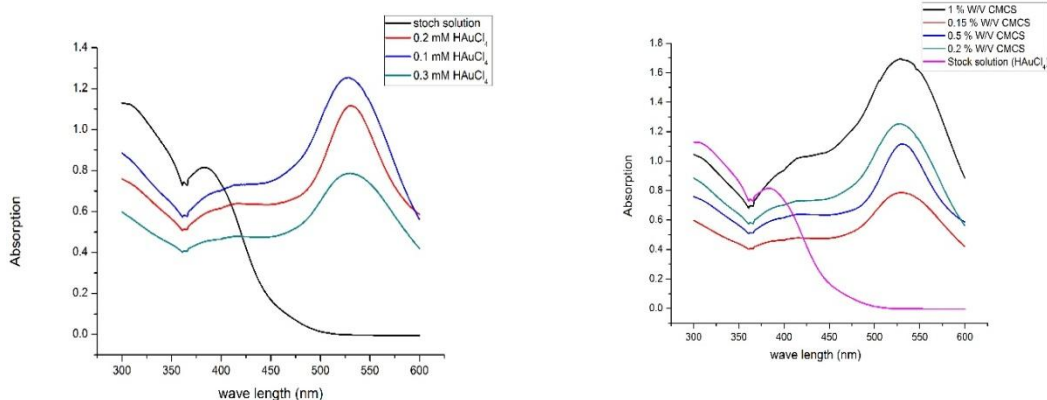
Figure 2 shows TEM images of the O-CMCS-AuNPs nanocomposites. It was observed that the AuNPs were encapsulated by O-CMCS and their size ranged from 15-25 nm. Unlike most gold colloid, few amount of non-spherical particles beside most spherical one was observed. These non-spherical particles seemed to be fabricated by gathered interference of two or three spherical particles during the nucleation process[14, 23].



**Figure 2:-**TEM images of O-CMCS-AuNPs nanocomposites.

Spectrophotometry is another important aspect for characterization of gold nanoparticles. With increase in particle size, the absorption peak shifts to longer wavelength and the width of absorption spectra is related to the size distribution range (Figure 2). Generally, gold nanospheres display a single absorption peak in the visible range between 510-550 nm, because of surface Plasmon resonance and show heavy absorption of visible light at 520 nm. This gives brilliant red color to Gold Nanoparticle (AuNPs), which varies according to their size. In present study

the absorption of gold nanoparticle was measured in single beam spectrophotometer and absorption maxima was noted at different wavelength (390-630 nm).



**Figure 3:-** UV/visible spectrum of gold nanoparticles

Figure 3 shows that the UV/visible spectrum of gold nanoparticles prepared by using carboxymethylchitosan as reducing agent as well as capping (stabilizing) agent with different gold salt concentrations (0.1, 0.2, 0.3 mM) and different O-CMCS concentration (0.15%, 0.2%, 0.5% and 1% W/V) to obtain the optimum condition and these results show that the optimum condition for preparing uniform AuNPs from O-CMCS at 0.2% CMCS and 0.2 mM HAuCl<sub>4</sub>.

#### Finishing of Fabrics with Gold Nanoparticles:-

We used 100% cotton (Sample 1) and 65:35 cotton: polyester (Sample 2) blend fabrics finished with the prepared gold nanoparticles (AuNPs) to be fabrics with new properties such as ultra violet protection (UPF). The results of the anti-UV efficacy of the untreated and AuNPs loaded substrates are shown in Table 1. It demonstrates that after treatment of fabrics with AuNPs results in a significant increase in their UV-protection function. The UV-protection property of the untreated cotton and Cotton/polyester substrates showed that they afforded poor protection, UPF < 20, against UV-radiation. The variation in protection value, expressed as UPF, between the cotton and Cotton/polyester, before and after post-treatment with AuNPs, is attributed to their differences in fabric construction [24].

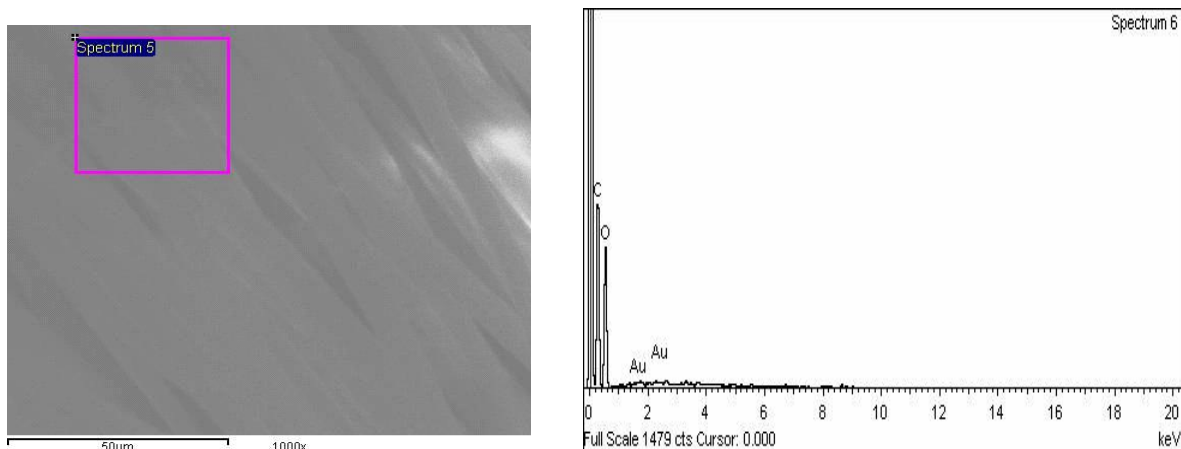
**Table 1:-** Atomic absorption and UPF values of the fabrics treated with AuNPs

Description	Atomic Absorption (mg/dl)	UPF
(Sample 1) untreated	0.000	8.211
(Sample 2) untreated	0.000	10.281
(Sample 1) treated with 15 mM AuNPs	1.203	20.54
(Sample 1) treated with 15 mM AuNPs	0.970	31.35
(Sample 1) treated with 20 mM AuNPs	1.493	36.12
(Sample 2) treated with 20 mM AuNPs	3.841	48.31
(Sample 2) treated with 25 mM AuNPs	3.841	50.65
(Sample 2) treated with 25 mM AuNPs	3.630	61.23

Scanning electron microscope (SEM/EDX) analysis were shown in Figures (4), which shows the presence of Au nanoparticles in the fabrics. The surface morphology of the treated fabric with Au nanoparticles appears as smooth surface with deposit of the nanoparticles. It is clear that the prepared Au nanoparticles are more homogenous and regular distribution on the surface and has higher intensity peaks, on the other hand the Au nanoparticles EDX analysis indicate that the content of Au (0.49, 0.29) (Au weight 0.91) and has lower intensity peaks [25].

The observation of the Au nanoparticles coating shows that the surface texture appears to have dense and low porosity (The choice samples were interlock). In case of ripe fabrics the nanoparticles coated the fibers and appears to be uniform in size.

The coated fabric with Au nanoparticles film was formed and firmly on the surface of the sample. It is evident that experimental and reaction conditions did not alter the morphology surface on using Au nanoparticles. The Au nanoparticles was strongly attached to the fibers due to very strong electrostatic or chemical interactions between the Au nanoparticles and the fabric [25].



**Figure 4:-SEM and EDX spectra cotton treated with AuNPs**

#### Cytotoxicity of GNPs suspensions:-

To study the effect of gold ions concentration present in AuNPs suspensions on their toxicity, A549 cells were treated for 24 h with three different batches of AuNPs suspension, which contained the same concentration of GNPs (1mM) in three GNP types. As shown in Table. Epiderm cell line selected for cytotoxicity test for Au NPs prepared from  $\text{HAuCl}_4$  solution.

#### Cytotoxicity evaluated using two protocols: EC50 and MTT:-

Table 2 shows that the  $\text{IC}_{50}$  of  $\text{HAuCl}_4$  (1 mM), gold nanoparticles prepared by citrate reduction and third with our method using carboxymethylchitosan (CMCS). However, its toxicity reduces in AuNPs treated with CMCS. In this study,  $\text{Au}^+$  decreased mitochondrial activity more than AuNPs with almost two fold difference in  $\text{IC}_{50}$  values as shown in Table 2, which agreed with previous studies of many researcher[25-28]

**Table 2:-** $\text{IC}_{50}$  A549 cell line after exposing to  $\text{HAuCl}_4$ , Au NPs (citrate reduction), Au NPs (CMCS reduction) (for 24 h)

Material	EC50,
$\text{HAuCl}_4$ (1 mM)	0.32µg/ml
Au NPs (citrate reduction)	0.21 µg/ml
Au NPs (CMCS reduction)	5.61 µg/ml

MTT assay used to measure the cell viability expressed in the decrease in mitochondrial activity (Table3). A reduction in mitochondrial function of A549 cells exposed to the three GNPs types prepared.

**Table 3:-**MTT Test (metabolic Activity of the Mitochondria):

Material	MTT expressed in viable cells	
	After 3hrs.	After 24 hrs.
$\text{HAuCl}_4$ (1 mM)	18.6	2.8
Au NPs (citrate reduction)	14.2	1.9
Au NPs (CMCS reduction)	84.5	76.2

#### Conclusion:-

- Gold nanoparticles (AuNPs) was prepared via simple and green method by using CMCS as reducing and stabilizing agents at the same time.
- CMCS prepared based on our previous method by reacting chitosan with monochloroacetic acid in alkaline medium.

- AuNPs were prepared by using different concentrations of carboxymethylchitosan (0.2% w/v, 0.5% w/v and 1% w/v) at 100 °C for 1 hour.
- CMCS was characterized by using nitrogen content, carboxyl content and FTIR spectra. AuNPs was characterized by using UV spectrophotometry and TEM images.
- The cytotoxicity of the prepared AuNPs were evaluated using cell viability assay from MMT and IC<sub>50</sub> values compared with AuNPs prepared by chemical methods. The results shows that AuNPs have normal distributed with 15-25 nm particle size and its cytotoxicity was lowered when prepared by this green method and can use GNPs safely in contact medical treatment with skin.

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